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Association of the Fatty Acid Amide Hydrolase C385A Polymorphism with Alcohol Use Severity and Coping Motives in Heavy-drinking Youth

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Conflict of Interest

RFT has consulted for Quinn Emanuel and Ethismos on unrelated topics. SJK receives funding from Jazz Pharmaceuticals and U.S. NIH NIAAA for studies unrelated to the present investigation. BLF has no relevant COI to declare. Unrelated to the present work: BLF has obtained funding from Pfizer (GRAND Awards, including salary support) for investigator-initiated projects. BLF has some in-kind donation of cannabis product from Aurora and medication donation from Pfizer and Bioprojet and was provided a coil for TMS study from Brainsway. BLF has obtained industry funding from Canopy (through research grants handled by CAMH or University of Toronto), Bioprojet, ACS and Alkermes. BLF has received in kind donations of nabiximols from GW Pharma for past studies funded by CIHR and NIH. All other authors declare no conflict of interest.

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Abstract

Background: Reduced function of fatty acid amide hydrolase, the catabolic enzyme for the endocannabinoid anandamide, can be inherited through a functional genetic polymorphism (*FAAH* rs324420, C385A, P129T). The minor (A) allele has been associated with reduced *FAAH* enzyme activity and increased risk for substance use disorders in adults. Whether this inherited difference in endocannabinoid metabolism relates to alcohol use disorder etiology and patterns of alcohol use in youth is unknown.

Methods: To examine this question, heavy-drinking youth ($n = 302$; mean age = 19.74 ± 1.18) were genotyped for *FAAH* C385A. All subjects completed a comprehensive interview assessing alcohol use patterns including the Timeline Follow-back Method, Alcohol Use Disorders Identification Test (AUDIT) and Drinking Motives Questionnaire. ANCOVAs were conducted to assess differences in drinking patterns and drinking motives between genotype groups, and mediation analyses investigated whether drinking motives accounted for indirect associations of genotype with alcohol use severity.

Results: Youth with the *FAAH* minor allele (AC or AA genotype) reported significantly more frequent drinking days ($p=0.045$), significantly more frequent heavy episodic drinking ($p=0.003$) and significantly higher consumption patterns (AUDIT $p=0.045$, AUDIT-C $p=0.02$). Mediation analyses suggested that the association of *FAAH* C385A with drinking outcomes was mediated by coping motives.

Conclusions: These findings extend previous studies by suggesting that reduced endocannabinoid metabolism may be related to heavier use of alcohol in youth, prior to the onset of chronic drinking problems. Furthermore, differences in negative reinforcement-related drinking might account in part for this association.

85

86 **KEY WORDS:** Alcohol Use Disorders Identification Test (AUDIT), Drinking Motives, Fatty

87 Acid Amide Hydrolase (FAAH), *FAAH* Polymorphism (C385A; rs324420), Youth

88

Introduction

Alcohol use is common in adolescence and young adulthood. Over 54% of college students in the United States ages 18-22 report alcohol consumption in the past month and 37% indicate at least one occasion of past-month heavy episodic drinking (HED: >4 and 5 drinks per occasion for females and males respectively) (National Institute on Alcohol Abuse and Alcoholism, 2020), a known early risk factor for subsequent development of alcohol use disorder (AUD) (Gowin et al., 2017, Greenfield et al., 2014, Hingson et al., 2017). The progression of occasional drinking to alcohol-related problems including AUD is influenced by multiple factors, including motives to drink (Grigsby et al., 2016), and it has been proposed that young adults are motivated to consume alcohol by the expected positive or negative reinforcement effects (Kuntsche et al., 2005). In particular, drinking to increase pleasure (enhancement motives) is related to higher rates of heavy drinking in youth, while drinking to decrease negative affective states such as anxiety or stress (coping motives) has been more closely linked with AUD and alcohol-related problems (Kuntsche et al., 2005, Neighbors et al., 2007, Hammarberg et al., 2017). Better understanding these motives may assist in investigating how “reward” and/or “relief” drinking might relate to genetic or biological risk factors for AUD.

Recent advancements in understanding AUD have implicated the endocannabinoid system. In particular, fatty acid amide hydrolase (FAAH), the enzyme primarily responsible for the degradation of one of the major endocannabinoid ligands, anandamide, may play a role in the biological vulnerability for AUD (Basavarajappa et al., 2019). In this regard, preclinical studies using FAAH knock-out and knock-in mice models with impaired anandamide degradation, and those using pharmacological inhibition of FAAH, have linked decreased FAAH activity with increased alcohol preference and intake, reduced ethanol sensitivity/intoxication, reduced

alcohol-induced locomotor impairment, and reduced withdrawal effects (Blednov et al., 2007, Zhou et al., 2016, Vinod et al., 2008, Basavarajappa et al., 2006, Zhou et al., 2017). These findings are in line with the observation that rats bred to be alcohol-preferring rats, such as AA Alko and sP Sardinian rats, have lower FAAH protein levels and enzymatic activity prior to alcohol exposure (Vinod et al., 2012, Hansson et al., 2007). Increased alcohol preference and reduced sensitivity to the effects of alcohol are believed to be mediated, at least in part, through the cannabinoid 1 (CB1) receptor, with the majority of preclinical studies suggesting that CB1 stimulation increases alcohol consumption, while CB1 receptor antagonism/inverse agonism decreases alcohol intake (Cippitelli et al., 2005, Linsenbardt and Boehm, 2009, Malinen and Hyytiä, 2008, Hansson et al., 2007, Colombo et al., 2002). It has therefore been proposed that reduced FAAH activity, resulting in increased anandamide, may influence alcohol intake via a CB1 receptor-mediated mechanism (Basavarajappa et al., 2019).

In humans, FAAH levels can vary due to a common missense single nucleotide polymorphism (SNP), *FAAH* rs324420 / C385A. This *FAAH* SNP consists of a cytosine-to-adenine substitution (C385A) resulting in the conversion of a conserved proline residue at amino acid position 129 to a threonine residue (P129T). Minor allele (A) homozygotes and heterozygotes (i.e., AA and AC genotypes, respectively), make up approximately 38% of individuals of European descent (Sipe et al., 2002). These individuals have reduced FAAH protein levels and steady-state enzymatic activity compared to CC homozygotes due to a post-translational mechanism (Chiang et al., 2004), which results in a higher concentration of anandamide (Mayo et al., 2018, Dincheva et al., 2015, Sipe et al., 2010, Boileau et al., 2015). Some (Sipe et al., 2002, Flanagan et al., 2006) but not all (Tyndale et al., 2007, Bühler et al., 2014) studies have shown that individuals with the minor (A) allele have increased vulnerability

for alcohol use and addiction. Of note, a recent study in adult humans of Caucasian/European ancestry with AUD reported that individuals with the AC or AA genotype engaged in significantly more drinking days and heavy drinking episodes and had greater self-reported alcohol use severity compared to CC homozygotes (Sloan et al., 2017). Consistent with this finding, low FAAH brain levels, as measured using positron emission tomography (PET), were associated with increased alcohol consumption in a sample of treatment-seeking individuals with AUD (Best et al., 2020). Together, these findings suggest that low FAAH activity may be related to increased alcohol consumption and vulnerability for AUD, though the mechanism by which this might occur requires further investigation.

In addition to alcohol consumption, FAAH is involved in responses to reward (Scherma et al., 2018), anxiety and stress (Gunduz-Cinar et al., 2013). Overexpression of FAAH attenuated stress response and decreased anxiety-like behaviour in one preclinical study (Morena et al., 2019). Other studies have found low FAAH, as indexed by *FAAH AC/AA genotype*, and, specifically, the *FAAH C385A* minor allele, to be related to lower anxiety and stress-relevant traits including enhanced fear extinction and fear extinction recall, as well as increased reward sensitivity, ventral striatal reactivity and impulsivity (Mayo et al., 2018, Hariri et al., 2009, Mayo et al., 2020). Further, some, though not all, studies of those with this variant have reported lower amygdala reactivity to threat during an fMRI task and self-reported anxiety (Dincheva et al., 2015, Hariri et al., 2009). These studies suggest that reduced FAAH activity may increase alcohol drinking and risk for AUD by impacting reward response and/or stress-reactivity, and that this relationship may therefore be mediated by positive or negative reinforcing motives for drinking. In particular, both positive reinforcement (enhancement) and negative reinforcement (coping) motives are associated with alcohol use severity, with coping motives being a particular

158 risk factor for alcohol-related problems and the development of AUD (Hammarberg et al., 2017,
159 Kuntsche et al., 2005, Galen et al., 2001, Mezquita et al., 2011). Deficient FAAH activity could
160 affect the experience of intoxication, rewarding properties of alcohol, and/or one's response to
161 stress, therefore affecting coping and/or enhancement motives for drinking. However, no
162 research to date has investigated the endocannabinoid system, and more specifically FAAH, in
163 relation drinking motives as risk factors in the development of AUD.

164 Another unanswered question is whether the association between low FAAH activity (as
165 indexed by AC/AA genotype) and increased alcohol consumption is evident only following the
166 emergence of AUD, or, is observable early in drinking history, potentially influencing the
167 development of alcohol use problems, as suggested by the preclinical literature. Further, there is
168 little known about the mechanisms by which the endocannabinoid system, including FAAH,
169 might relate to motivation to consume alcohol and the risk for developing AUD. As such, the
170 present study sought to investigate whether the FAAH C385A minor (A) allele (i.e., AC or AA
171 genotype) relates to alcohol consumption and drinking motives in youth. Based on the preclinical
172 literature related to the endocannabinoid system and findings in humans with AUD, we
173 hypothesized that youth with the AC or AA genotype (i.e., those presumed to have lower FAAH
174 activity), would exhibit more hazardous patterns of alcohol consumption as compared to CC
175 homozygotes, and that this relationship may be mediated by genotype differences in coping or
176 enhancement drinking motives.

177 178 **Materials and Methods**

179 **Participants and recruitment**

Participants were recruited from community and University settings in Toronto, Canada for participation in one of two larger studies investigating the risk for AUD in youth (Hendershot et al., 2017, Best et al., 2020). To be eligible for inclusion in either of the two larger studies, participants were at least 18 years of age, reported at least one occurrence of HED in the previous 30 days, no current medication or medical condition contraindicating alcohol consumption and no current diagnosis or treatment of a psychiatric condition or substance use disorder, as assessed by the Structured Clinical Interview for DSM-IV (SCID-IV) or the Mini International Neuropsychiatric Interview (MINI). Participants reported no current or past attempts to seek treatment for drinking, and no current or past history of significant alcohol related problems, based on telephone screening questions and the brief Michigan Alcohol Screening Test (Pokorny et al., 1972). Age was restricted in one of the two studies to the range of 18-21 years, while the other study included participants aged 19-25. Regular, recreational cannabis use was not an exclusion criterion for one study but was exclusionary for the second. Demographic information by parent study are reported in supplementary material ([Supplementary Table 1](#)). The total sample for the current analyses (N=302) comprised all participants with valid genotype data who also had complete data on the Timeline Follow Back (TLFB) and Alcohol Use Disorders Identification Test (AUDIT) questionnaire. Forty additional participants were consented in one of the studies but were not included in the current analyses due to missing genotype data (n = 18) and/or incomplete questionnaires (AUDIT n = 7, TLFB n = 15).

Demographics are reported in [Table 1](#). The sample was comprised of 53% women. 51% identified as Caucasian/of European descent and 78% were currently enrolled as post-secondary students. All participants were 19-25 years of age (mean: 19.88 ± 1.2) and reported at least one

heavy drinking episode in the past month. Alcohol use, AUDIT and Drinking Motive Questionnaire Coping and Enhancement scores are reported in Table 2.

Procedures

All procedures were approved by the Centre for Addiction and Mental Health Research Ethics Board. All participants completed an initial phone screening questionnaire; those who met age and alcohol consumption criteria (at least one occasion of HED in the past month) without any current, serious pre-existing medical, psychiatric or substance use conditions were invited for a baseline session. Data were collected during the baseline session prior to the completion of any other laboratory and/or neuroimaging sessions that the participant may have qualified for. Questionnaires were administered to assess recent alcohol use (described below). In addition, recent cannabis use was assessed and the Barratt Impulsivity Scale (BIS) was used to evaluate impulsivity. Urine samples were collected to test for recent use of other drugs of abuse.

Measures

Alcohol Consumption. A Timeline Follow Back (TLFB) interview (Sobell and Sobell, 1992) was used to assess alcohol use over the past 90 days. The TLFB is a calendar-based assessment that uses cues to facilitate accurate recall of recent drinking behavior. Participants reported the number of standard drinks they consumed on each day over the past 90 days, and data were used to derive indices of past 90-day drinking frequency, average number of drinks consumed per drinking occasion, and frequency of heavy episodic drinking (defined as 4/5 or more drinks per occasion for women/men). Participants completed the 10-item AUDIT (Saunders et al., 1993), which contains items assessing alcohol consumption, alcohol-related

harms, and alcohol dependence symptoms. The AUDIT-C (comprising the AUDIT consumption items, items 1-3, range: 0-12) provides an index of consumption, whereas the AUDIT total score (range: 0-40) provides a continuous index of hazardous drinking severity.

Drinking Motives. Two 5-item subscales of the Drinking Motives Questionnaire-Revised (Cooper, 1994) were used to assess internal reinforcement motives for positive reinforcement and negative reinforcement drinking (i.e., enhancement and coping motives). The enhancement motives scale is comprised of the sum of 5 items assessing the frequency of drinking for the enjoyment or pleasure derived from alcohol consumption (e.g. “Because it gives you a pleasant feeling”). The coping motives scale is, likewise, comprised of the sum of 5 items assessing the frequency of drinking to reduce negative mood and to help put worries out of one’s mind (e.g., “To forget about your problems”). Participants rated each item on a scale from 1 = *almost never/never* to 5 = *almost always/always*. Other drinking motives domains (e.g., social and conformity motives) were not examined because our hypotheses concerned *FAAH* genotype differences in internal (rather than external/social) aspects of alcohol reward or reinforcement.

Cannabis and Nicotine Use. The NIDA-Modified Alcohol Smoking and Substance Involvement Screening Test (NM ASSIST) was used to assess recent, self-reported cannabis use frequency over the last 90 days. Participants responded on a 5-point scale ranging from *Never* to *Daily or Almost Daily*. Participants recruited through the smaller of the two parent studies reported their recent use of cannabis using the 90 day TLFB, if applicable. From this, the number of days per week on which cannabis was used was determined and this data was adapted to the NM ASSIST. For example, participants reporting no occasions of cannabis use in the past ninety days were scored “*Never*” on the NM ASSIST, whereas participants reporting one or two occasions of cannabis use on the 90-day TLFB were scored as “*once or twice*”. Those reporting

weekly cannabis use less than once per week, but at least once per month were scored on the NM ASSIST as “*monthly*”. Finally, one participant reported using cannabis once per week, and was scored “*weekly*” on the NM ASSIST. It should be noted that positive cannabis urine toxicology was an exclusion criteria for this smaller study, and, as such, these participants reported minimal cannabis use (see Supplementary Table 1 for more details). Nicotine use was self-reported, and nicotine dependence was assessed using the Fagerström test of Nicotine Dependence (Heatherton et al., 1991).

Genotyping. DNA was extracted from either venous blood samples (n = 23) or saliva samples (n = 279); a 260/280 ratio of above 1.8 was the minimal quality for DNA used. *FAAH* genotyping (rs324420) was performed as previously described (Boileau et al., 2015). Briefly, the *FAAH* genotype (rs324420) was determined using the Taqman SNP genotyping assay set performed on a ViiA7 thermal cycler (Life Technologies, Burlington, Ontario, Canada) with appropriate controls. Five µl of 2x GTXpress Master mix (cat#4401892, Life Technologies) was mixed with 10 ng of DNA and the 40 × probe (cat#C_1897306_10, Life Technologies) in a final volume of 10 µl and run for 50 cycles of 95°C for 1 second and 60°C for 20 seconds. Positive controls include DNA samples of all three genotypes and negative controls include samples with DNA replaced by water; a minimal ViiA7 quality score of 98% was required.

Data Analysis

Prior to analyses, distributions of all variables were screened for extreme outliers (defined as greater than 3.29 standard deviations from the mean and being notably disconnected from the other scores (Tabachnick et al., 2007). Extreme outliers were observed on the drinking frequency (n=1) and heavy drinking frequency (n=2) variables; these values were Winsorized

(i.e., recoded to one unit greater than the next most extreme value) to reduce their influence in the models. We then examined descriptive statistics comparing *FAAH* genotype groups on demographic variables. Given that only $n=14$ participants were homozygous for the A allele, we had low power to examine these participants separately in the main analyses. Thus, we combined participants that were AA homozygous with those AC heterozygous into an AC/AA, genotype group (*AC/AA group*) ($n=116$) and compared them to participants homozygous for the C allele (*CC group*) in all analyses (Sloan et al., 2017, Dincheva et al., 2015). Differences in demographic characteristics, including cannabis use and BIS impulsivity scores, were assessed using independent samples t-tests or chi-squared tests, where applicable.

In order to test the hypothesis that the AC/AA group would drink more heavily and hazardously relative to the CC group, we conducted a series of ANCOVAs comparing frequency of drinking, average drinks per occasion, frequency of heavy episodic drinking, and total AUDIT score between the AC/AA group and CC group. We also ran an ANCOVA with enhancement and coping motives as outcomes to explore whether there were genotype differences in coping-related drinking motivations. ANCOVAs included participant sex and race (white vs. non-white) and age as covariates. We also examined models including Genotype*Sex and Genotype*Race interactions to explore whether sex and race moderated any effects of genotype on drinking outcomes. Although the original studies were not powered based on this research aim, the sample size allowed for detection of effect sizes as small as partial $\eta^2 = 0.026$ for the covariate adjusted mean differences between *FAAH* genotype groups with 80% power and alpha of 0.05. For comparison, the current study was powered to detect standardized differences in unadjusted means as small as $d = 0.33$, which is on the lower end of the range reported in the largest study on *FAAH* and alcohol to date, which found genotype differences in AUDIT scores with effect

sizes ranging from $d = 0.26$ (African American sample, $n = 144$) to $d = 0.74$ (Caucasian sample, $n = 63$).

Next, we tested the hypothesis that coping or enhancement drinking motives might mediate the relationship between genotype and drinking outcomes. To do so, for any motive subscale that was significantly different across genotype groups (based on the ANCOVAs described above), its role as a mediator was examined using the SPSS PROCESS macro, which uses bootstrapping to derive bias corrected confidence intervals (CI) of the mediated effect (Hayes, 2017). In each model, we specified sequential indirect effect pathways from genotype to coping motives to alcohol consumption to AUDIT. Specifically, genotype group ($CC=0$, AC or $AA=1$) was specified as a predictor of coping or enhancement motives (z -score transformed), which in turn was specified as a predictor of each alcohol consumption index (drinking frequency, average drinks per occasion, heavy episodic drinking frequency, each examined in a separate model). Given that heavier alcohol consumption is thought to underlie hazardous/harmful drinking outcomes, each model then included hazardous drinking severity assessed with the AUDIT as the final outcome. For all mediation models, sex, race and age were included as covariates in all estimated paths. Support for significant mediation was observed when the 95% bias corrected confidence interval for the indirect effect did not contain zero.

SPSS Statistics software was used for all analyses, and level of significance was set at $p < 0.05$.

Results

[Table 1](#) shows participant demographics by genotype group. The groups did not differ significantly with respect to sex, ethnicity and proportion of currently enrolled students. Of the 302 participants, 61.6% were CC homozygotes, 33.8% AC heterozygotes and 4.6% AA

homozygotes. The minor (A) allele frequency in this sample (.215) was consistent with other reports involving predominantly Caucasian participants (e.g., Sloan et al., 2017), and genotype distributions did not diverge from Hardy-Weinberg equilibrium. The AC/AA group had a significantly higher mean age than the CC genotype group, though all enrolled participants were between 18 and 25 years of age. Scores on the Barratt Impulsivity Scale did not significantly differ between genotype groups. Further, we found no significant difference in frequency of smokers, level of tobacco dependence or cannabis use between groups.

Table 2 shows the covariate-adjusted means, standard deviations, F tests, and effect size estimates (partial η^2) from the ANCOVAs comparing the AC/AA and CC groups on alcohol-related outcomes and enhancement and coping drinking motives (controlling for sex, race, and age). Supplementary Table 2 contains the unadjusted means, standard deviations and standardized mean differences (d) between the genotype groups on each of the outcomes. As shown in Table 2, the AC/AA group reported significantly greater frequency of alcohol consumption and significantly greater heavy episodic drinking frequency, without a significant difference in the average number of drinks per occasion (Figure 1a). The AC/AA group also had a significantly higher total AUDIT scores (Figure 1a) than the CC group (Table 2). AUDIT-C (consumption) scores were also significantly elevated in AC/AA group (Figure 1a) compared to the CC group. Finally, the AC/AA group had significantly greater endorsement of coping motives for drinking (Figure 1b), but no difference in enhancement drinking motives (Figure 1b). Effect sizes for all of the significant group differences were small (partial η^2 ranged from .013 to .029), suggesting that that genotype group uniquely accounted for 1%-3% of the variance in these outcomes. Of note, none of the interactions between genotype and sex or genotype and race were significant in any of these models (all $p > .05$), suggesting that sex and race did not moderate

the effects of genotype. Further, when including sample source (i.e., which of the two studies the participants were drawn from) as an additional covariate in the ANCOVAs, none of the main effects for genotype changed, and there were no main effects of sample source on any of the outcomes (all $p > 0.21$).

Significant main effects of sex were observed for the AUDIT, AUDIT-C, and drinks per occasion (all $p < .05$), with men having higher means than women. Significant main effects of race were observed for AUDIT-C, drinking frequency, and heavy episodic drinking frequency (all $p < .05$), with white participants having higher means than non-white participants. Age was also a significant covariate in several models, with older participants reporting more frequent drinking, lower average drinks per occasion, lower AUDIT total scores, and lower coping and enhancement motives (all $p < .05$).

Next, coping motives were investigated as a potential mediator of the relationship between genotype and each alcohol consumption outcome (frequency, quantity, heavy drinking episode), which in turn were specified as mediators of the association between coping motives and AUDIT score. As enhancement motives did not differ between genotype groups, only coping motives were tested in these mediation analyses. The results of these analyses are depicted in [Figure 2](#). In all models, genotype was a significant predictor of coping motives, and coping motives, in turn, significantly predicted the alcohol outcomes. The alcohol outcomes, in turn, predicted AUDIT scores in all three models. In all models, the indirect association from genotype to coping motives to alcohol consumption (frequency, quantity, or heavy episodic drinking) to AUDIT was statistically significant as indicated by 95% CIs that did not contain zero. In addition, in the models that included drinking frequency and average drinks per occasion, the direct paths from genotype to the drinking variable was not significant when

controlling for the mediated pathway through coping motives (see Figure 2a and 2b), suggesting that coping motives fully mediated the pathway from genotype to alcohol use and AUDIT in these models. Further, in all models, the direct path from coping motives to AUDIT was statistically significant, suggesting that alcohol consumption variables only partially mediated the link between coping motives and AUDIT in these models.

Discussion

The present study reports that youth with the A allele (AC or AA genotype) of the *FAAH* C385A polymorphism (i.e., those with presumed reductions in endogenous FAAH) engage in more hazardous drinking patterns through more frequent HED and reporting higher AUDIT scores. This group also reported significantly higher endorsement of coping motives for drinking. Further, our mediation models suggest that there is a significant indirect effect of genotype with risk for more hazardous drinking, such that the AC/AA group has a greater tendency to drink for coping-related reasons, which in turn relates to more frequent and heavier alcohol consumption, and more hazardous drinking outcomes (AUDIT).

Our findings support and extend the current literature in which lower FAAH, through genotypic differences or pharmacological manipulation, has been associated with AUD (Sloan et al., 2017, Best et al., 2020), increased severity of alcohol consumption (Basavarajappa et al., 2019). The present study reports complementary data to that which has incorporated neuroimaging and blood phenotypes to investigate the endocannabinoid system in relation to AUD and risk for addiction (Best et al., 2020, Boileau et al., 2016, Mansouri et al., 2020). As previous studies have focused on preclinical and/or adult populations, the present study is the second (and largest to date) to investigate this relationship in a youth population. Consistent with our findings, Sloan *et al.* reported that older Caucasian adults (mean age of 39 years) with the

AC/AA genotype with AUD had a significantly higher frequency of heavy drinking episodes and higher AUDIT scores compared to CC homozygotes (Sloan et al., 2017). As expected, based on the difference in populations investigated, Sloan *et al.*, reported higher rates of alcohol use compared to the present study. Effect sizes for genotype differences in drinking outcomes were in the small range in the present study (Table 2), which is consistent with the lower end of the range of effect sizes reported in Sloan *et al.* (see also Supplementary Table 2 for comparison), and would be expected in genetic association studies. As such, any clinical significance of these findings remains unclear. Nonetheless, our finding of a mean difference in AUDIT scores of 1.33 could imply clinical significance in some contexts, given that a 1-point difference in AUDIT score could reflect changes in consumption of 1-2 drinks per day or up to 30 drinks per week, depending on the item. Together, these two studies represent the largest samples investigated to date and suggest an association between AC/AA genotypes for the *FAAH* polymorphism and increased vulnerability for more hazardous alcohol consumption that persists across age and diagnostic groups.

By focusing on a youth sample, the current study begins to translate to humans the preclinical literature in which alcohol-naïve alcohol-preferring rats were found to have low *FAAH* (Vinod et al., 2012, Hansson et al., 2007) and genetically engineered mice homozygous for the A-allele of the *FAAH* C385A SNP showed increased “binge” alcohol consumption (Zhou et al., 2016). Our finding is, however, inconsistent with another smaller study in youth which used a different method of measuring ‘risky drinking’ (Bühler et al., 2014), as compared to the 90 day TLFB used in the present and other related study (Sloan et al., 2017). By studying both alcohol consumption and drinking motives in a large sample of young drinkers, the present study is the first to investigate in humans whether the relationship of low *FAAH* activity (due to the

FAAH C385A minor allele) and increased hazardous alcohol consumption is relevant to AUD etiology, and specifically, whether the association between *FAAH* genotype and drinking motives reflects a possible mechanism.

Here we report the association of inherited low *FAAH* activity with increased hazardous alcohol consumption and drinking motives, specifically negative reinforcement/coping motives (i.e., drinking to reduce negative affect); we found no association with positive reinforcement/enhancement motives (i.e., drinking to enhance positive affect) or impulsivity.

There are multiple ways that the presence of this *FAAH* C385A polymorphism, which has been shown to decrease *FAAH* activity in brain and increase anandamide across species (Dincheva et al., 2015, Mayo et al., 2018, Boileau et al., 2015, Spagnolo et al., 2016, Sipe et al., 2010), might increase alcohol consumption. One possibility is that impaired anandamide degradation may increase the rewarding properties of alcohol through the established, yet complex interaction between endocannabinoids and the dopaminergic system. In relation to the *FAAH* polymorphism, heterozygous AC mice have enhanced mesolimbic dopamine circuitry (Burgdorf et al., 2020), while humans and mice with the *FAAH* C385A polymorphism have significantly elevated D3 receptor levels in the brain (Mansouri et al., 2020), and increased striatal reward reactivity (Hariri et al., 2009). This suggests that dysregulation of the endocannabinoid system might impact the dopaminergic response to reward, in turn driving drug-seeking behavior.

In addition, alcohol's rewarding or negative reinforcement properties may also be enhanced through functional interactions of the endocannabinoid and opioid systems (Haller et al., 2008), or through modulation of alcohol-induced GABAergic neurotransmission. As indicated by studies in *FAAH* KO mice, low *FAAH* increased ethanol-reward behavior, decreased sensitivity to ethanol-induced effects and reduced mu-opioid activation as compared to

wild-type animals (Vinod et al., 2008). It has also been suggested, based on studies in mouse models, that decreased FAAH activity results in reduced sensitivity to alcohol's effects, including intoxication and sedation, and that differences in GABAergic neurotransmission may contribute to these effects (Pavon et al., 2019, Zhou et al., 2016). In humans, this could translate to differential sensitivity to acute alcohol effects, which in turn might influence alcohol consumption. In particular, our finding that inherited low FAAH activity relates specifically to coping, but not enhancement, motives, raises the possibility that individuals with lower FAAH may show differences in the magnitude of sedative or anxiolytic effects experienced under acute alcohol. An alternate possibility is that sedative or anxiolytic effects of alcohol do not differ in magnitude but are nonetheless experienced as more rewarding for these individuals. Either possibility could promote greater use of alcohol for negative reinforcement reasons (i.e. drug taking that alleviates a negative emotional state (Koob, 2014)).

The current study is the first to report coping motives as a mediator of the previously reported finding that individuals with the *FAAH* C385A minor allele report increased frequency and more hazardous patterns of alcohol consumption (Sloan et al., 2017). While this suggests that reduced FAAH activity may influence increased alcohol consumption in part through negative reinforcement mechanisms, the association between FAAH and coping motives is complex and there is limited literature exploring the endocannabinoid system, coping and drinking motives, specifically. In line with our findings, it has been established that youth who endorse drinking to cope report increased alcohol consumption and have increased risk for negative outcomes related to alcohol (Corbin et al., 2013, Grigsby et al., 2016). However, the preclinical and clinical literature generally suggests that lower FAAH is associated with reduced anxiety during alcohol withdrawal (Cippitelli et al., 2008), lower trait anxiety, reduced amygdala

456 reactivity to threats and lower stress reactivity (Dincheva et al., 2015, Mayo et al., 2018, Hariri et
457 al., 2009, Patel et al., 2017). Acute and chronic stress has been shown to upregulate FAAH
458 protein expression, such that individuals with the AC or AA genotype (Gray et al., 2015, Gray et
459 al., 2016), those with less functional FAAH protein, would catalyze less anandamide than CC
460 homozygotes under conditions of stress. Furthermore, endocannabinoid system activation via
461 pharmacological FAAH inhibition has been shown to attenuate stress-induced hypothalamic-
462 pituitary-adrenal axis activity (Gray et al., 2015).

463 Given evidence that a genetic predisposition to lower FAAH activity generally relates to
464 lower anxiety/stress reactivity, the current evidence for an association with *greater* motivation
465 for negative reinforcement drinking may at first appear contradictory. However, the anxiolytic
466 effects of decreased FAAH activity are notably more prominent under conditions of high
467 environmental aversiveness (Patel et al., 2017) and FAAH inhibition has also been shown to
468 attenuate stress-induced reductions in salience and reward-seeking behavior (Rademacher et al.,
469 2008). Further, FAAH and endocannabinoid tone may influence coping behavior such that
470 FAAH inhibition and increased anandamide promote more focused problem solving in light of
471 anxiety-inducing environmental stimuli (Aliczki et al., 2016). Finally, alcohol-related stress-
472 response has been shown to be context-dependent, whereby acute alcohol intoxication dampens
473 aversive stress-reactivity to uncertain or variable threat (Gorka et al., 2020, Bradford et al.,
474 2013). Together with the decreased experience of alcohol intoxicating effects seen in preclinical
475 investigations, individuals with the AC or AA genotype may therefore be inclined to consume
476 alcohol for the alleviation of negative affect in a context-dependent manner. Though negative
477 reinforcement is traditionally associated with later stages of AUD development, the present
478 relationship with coping motives presents a potential mechanism by which those with the *FAAH*

minor allele may be at greater risk for developing AUD, but future research is necessary to further elucidate these relationships.

Our study is not without limitations. While other studies investigating the *FAAH* C385A polymorphism and addiction or alcohol use have found ethnic-specific effects, our limited sample size did not allow for investigation of the impact of *FAAH* genotype on alcohol consumption patterns within an ethnic group. One previous study found an association of the AA homozygous genotype with alcohol abuse and addiction in a predominantly African American group (Flanagan et al., 2006), while another only found an effect of the A-allele in Caucasian individuals with AUD (Sloan et al., 2017). Though ethnicity was taken into consideration as a covariate, it is difficult to predict how this may have impacted our results. Another limitation of the present and other clinical studies investigating *FAAH* and AUD is that alcohol consumption data have been based on self-report using measures such as the AUDIT or TLFB. To more thoroughly investigate this relationship and support the translation of findings from alcohol-administration procedures in preclinical studies, laboratory-based alcohol challenge studies are warranted in humans.

Other limitations of the current study include the cross-sectional design and the inability to examine individuals with the AA homozygote genotype independently, as we may not be able to account for differential effects in this specific genotype, as some studies have reported. We acknowledge that there has been limited assessment of other relevant phenotypes, including anxiety, and as such, cannot rule out the potential impact of this on our results. Additionally, comprehensive diagnostic information was not collected on all participants included in the present analyses, therefore it cannot be ruled out that some participants may have reported early symptoms of AUD. Finally, the present study did not directly measure *FAAH* or *FAAH* substrate

502 concentrations (e.g. anandamide, oleoylethanolamide or palmitoylethanolamide). It cannot be
503 ruled out that the presently reported association between the *FAAH* polymorphism and increased
504 alcohol consumption may also be influenced by increased FAAH substrates other than
505 anandamide (e.g. oleoylethanolamide and palmitoylethanolamide) and future studies would
506 benefit from these measurements in addition to genotype. Nonetheless, the present study
507 provides a developmental context for the investigation of the impact of FAAH on AUD by
508 suggesting that innate differences in the endocannabinoid system relate to differences in alcohol
509 consumption patterns in youth, potentially via differences in the negative reinforcement value of
510 alcohol consumption. The association of low FAAH and increased risk for the development of
511 AUD may indeed predate the onset of alcohol problems, and more research is warranted to
512 further understand the mediators of this relationship.

References

- ALICZKI, M., BARNA, I., TILL, I., BARANYI, M., SPERLAGH, B., GOLDBERG, S. R. & HALLER, J. 2016. The effects anandamide signaling in the prelimbic cortex and basolateral amygdala on coping with environmental stimuli in rats. *Psychopharmacology (Berl)*, 233, 1889-99.
- BASAVARAJAPPA, B. S., JOSHI, V., SHIVAKUMAR, M. & SUBBANNA, S. 2019. Distinct Functions of Endogenous Cannabinoid System in Alcohol Abuse Disorders. *Br J Pharmacol*, [Epub ahead of print], 3085-3109.
- BASAVARAJAPPA, B. S., YALAMANCHILI, R., CRAVATT, B. F., COOPER, T. B. & HUNGUND, B. L. 2006. Increased ethanol consumption and preference and decreased ethanol sensitivity in female FAAH knockout mice. *Neuropharmacology*, 50, 834-844.
- BEST, L. M., WILLIAMS, B., LE FOLL, B., MANSOURI, E., BAZINET, R. P., LIN, L., DE LUCA, V., LAGZDINS, D., RUSJAN, P., TYNDALE, R. F., WILSON, A. A., HENDERSHOT, C. S., HEILIG, M., HOULE, S., TONG, J., KISH, S. J. & BOILEAU, I. 2020. Lower brain fatty acid amide hydrolase in treatment-seeking patients with alcohol use disorder: a positron emission tomography study with [C-11]CURB. *Neuropsychopharmacology*.
- BLEDNOV, Y. A., CRAVATT, B. F., BOEHM, S. L., WALKER, D. & HARRIS, R. A. 2007. Role of endocannabinoids in alcohol consumption and intoxication: studies of mice lacking fatty acid amide hydrolase. *Neuropsychopharmacology*, 32, 1570-1582.
- BOILEAU, I., MANSOURI, E., WILLIAMS, B., LE FOLL, B., RUSJAN, P., MIZRAHI, R., TYNDALE, R. F., HUESTIS, M. A., PAYER, D. E., WILSON, A. A., HOULE, S., KISH, S. J. & TONG, J. 2016. Fatty

534 Acid Amide Hydrolase Binding in Brain of Cannabis Users: Imaging With the Novel
535 Radiotracer [11C]CURB. *Biological psychiatry*, 80, 691-701.

536 BOILEAU, I., TYNDALE, R. F., WILLIAMS, B., MANSOURI, E., WESTWOOD, D. J., LE FOLL, B., RUSJAN,
537 P., MIZRAHI, R., DE LUCA, V., ZHOU, Q., WILSON, A. A., HOULE, S., KISH, S. J. & TONG, J.
538 2015. The fatty acid amide hydrolase C385A variant affects brain binding of the positron
539 emission tomography tracer [11C]CURB. *J Cereb Blood Flow Metab*, 35, 1237-1240.

540 BRADFORD, D. E., SHAPIRO, B. L. & CURTIN, J. J. 2013. How bad could it be? Alcohol dampens
541 stress responses to threat of uncertain intensity. *Psychol Sci*, 24, 2541-9.

542 BÜHLER, K. M., HUERTAS, E., ECHEVERRY-ALZATE, V., GINÉ, E., MOLTÓ, E., MONTOLIU, L. &
543 LÓPEZ-MORENO, J. A. 2014. Risky alcohol consumption in young people is associated with
544 the fatty acid amide hydrolase gene polymorphism C385A and affective rating of drug
545 pictures. *Mol Genet Genomics*, 289, 279–289.

546 BURGDORF, C. E., JING, D., YANG, R., HUANG, C., HILL, M. N., MACKIE, K., MILNER, T. A., PICKEL,
547 V. M., LEE, F. S. & RAJADHYAKSHA, A. M. 2020. Endocannabinoid genetic variation
548 enhances vulnerability to THC reward in adolescent female mice. *Sci Adv*, 6, eaay1502.

549 CHIANG, K., GERBER, A. L., SIPE, J. C. & CRAVATT, B. F. 2004. Reduced cellular expression and
550 activity of the P129T mutant of human fatty acid amide hydrolase: evidence for a link
551 between defects in the endocannabinoid system and problem drug use. *Human*
552 *Molecular Genetics*, 13, 2113-2119.

553 CIPPITELLI, A., BILBAO, A., HANSSON, A. C., DEL ARCO, I., SOMMER, W., HEILIG, M., MASSI, M.,
554 BERMÚDEZ-SILVA, F. J., NAVARRO, M., CICCOCIOPPPO, R. & DE FONSECA, F. R. 2005.

555 Cannabinoid CB1 receptor antagonism reduces conditioned reinstatement of ethanol-
556 seeking behavior in rats. *Eur J Neurosci*, 21, 2243-51.

557 CIPPITELLI, A., CANNELLA, N., BRACONI, S., DURANTI, A., TONTINI, A., BILBAO, A., RODRIGUEZ DE
558 FONSECA, F., PIOMELLI, D. & CICCOCIOPPO, R. 2008. Increase of brain endocannabinoid
559 anandamide levels by FAAH inhibition and alcohol abuse behaviours in the rat.
560 *Psychopharmacology*, 198, 449-60.

561 COLOMBO, G., SERRA, S., BRUNETTI, G., GOMEZ, R., MELIS, S., VACCA, G., CARAI, M. M. & GESSA,
562 L. 2002. Stimulation of voluntary ethanol intake by cannabinoid receptor agonists in
563 ethanol-preferring sP rats. *Psychopharmacology (Berl)*, 159, 181-7.

564 COOPER, M. L. 1994. Motivations for Alcohol Use Among Adolescents: Development and
565 Validation of a Four-Factor Model. *Psychological Assessment*, 6, 117-28.

566 CORBIN, W. R., FARMER, N. M. & NOLEN-HOEKESMA, S. 2013. Relations among stress, coping
567 strategies, coping motives, alcohol consumption and related problems: a mediated
568 moderation model. *Addict Behav*, 38, 1912-9.

569 DINCHEVA, I., DRYSDALE, A. T., HARTLEY, C. A., JOHNSON, D. C., JING, D., KING, E. C., RA, S., GRAY,
570 J. M., YANG, R., DEGRUCCIO, A. M., HUANG, C., CRAVATT, B. F., GLATT, C. E., HILL, M. N.,
571 CASEY, B. J. & LEE, F. S. 2015. FAAH genetic variation enhances fronto-amygdala function
572 in mouse and human. *Nat Commun*, 6, 6395.

573 FLANAGAN, J. M., GERBER, A. L., CADET, J. L., BEUTLER, E. & SIPE, J. C. 2006. The fatty acid amide
574 hydrolase 385 A/A (P129T) variant: haplotype analysis of an ancient missense mutation
575 and validation of risk for drug addiction. *Human genetics*, 120, 581-588.

576 GALEN, L. W., HENDERSON, M. J. & COOVERT, M. D. 2001. Alcohol expectancies and motives in a
577 substance abusing male treatment sample. *Drug Alcohol Depend*, 62, 205-14.

578 GORKA, S. M., KREUTZER, K. A., PETREY, K. M., RADOMAN, M. & PHAN, K. L. 2020. Behavioral and
579 neural sensitivity to uncertain threat in individuals with alcohol use disorder: Associations
580 with drinking behaviors and motives. *Addict Biol*, 25, e12774.

581 GOWIN, J. L., SLOAN, M. E., STANGL, B. L., VATSALYA, V. & RAMCHANDANI, V. A. 2017.
582 Vulnerability for Alcohol Use Disorder and Rate of Alcohol Consumption. *Am J Psychiatry*,
583 174, 1094-1101.

584 GRAY, J. M., VECCHIARELLI, H. A., MORENA, M., LEE, T. T., HERMANSON, D. J., KIM, A. B.,
585 MCLAUGHLIN, R. J., HASSAN, K. I., KÜHNE, C., WOTJAK, C. T., DEUSSING, J. M., PATEL, S.
586 & HILL, M. N. 2015. Corticotropin-releasing hormone drives anandamide hydrolysis in the
587 amygdala to promote anxiety. *J Neurosci*, 35, 3879-92.

588 GRAY, J. M., WILSON, C. D., LEE, T. T. Y., PITTMAN, Q. J., DEUSSING, J. M., HILLARD, C. J., MCEWEN,
589 B. S., SCHULKIN, J., KARATSOREOS, I. N., PATEL, S. & HILL, M. N. 2016. Sustained
590 Glucocorticoid Exposure Recruits Cortico-limbic CRH Signaling to Modulate
591 Endocannabinoid Function. *Psychoneuroendocrinology*, 66, 151-58.

592 GREENFIELD, T. K., YE, Y., BOND, J., KERR, W. C., NAYAK, M. B., KASKUTAS, L. A., ANTON, R. F.,
593 LITTEN, R. Z. & KRANZLER, H. R. 2014. Risks of alcohol use disorders related to drinking
594 patterns in the U.S. general population. *J Stud Alcohol Drugs*, 75, 319-27.

595 GRIGSBY, T. J., FORSTER, M., UNGER, J. B. & SUSSMAN, S. 2016. Predictors of alcohol-related
596 negative consequences in adolescents: A systematic review of the literature and
597 implications for future research. *J Adolesc*, 48, 18-35.

598 GUNDUZ-CINAR, O., HILL, M. N., MCEWEN, B. S. & HOLMES, A. 2013. Amygdala FAAH and
 599 anandamide: mediating protection and recovery from stress. *2013*, 34, 637-644.

600 HALLER, V. L., STEVENS, D. L. & WELCH, S. P. 2008. Modulation of opioids via protection of
 601 anandamide degradation by fatty acid amide hydrolase. *Eur J Pharmacol*, 600, 50-8.

602 HAMMARBERG, A., ÖSTER, C. & NEHLIN, C. 2017. Drinking motives of adult patients seeking
 603 treatment for problematic alcohol use. *J Addict Dis*, 36, 127-135.

604 HANSSON, A. C., BERMÚDEZ-SILVA, F. J., MALINEN, H., HYYTIÄ, P., SANCHEZ-VERA, I., RIMONDINI,
 605 R., RODRIGUEZ DE FONSECA, F., KUNOS, G., SOMMER, W. H. & HEILIG, M. 2007. Genetic
 606 impairment of frontocortical endocannabinoid degradation and high alcohol preference.
 607 *Neuropsychopharmacology*, 32, 117-126.

608 HARIRI, A. R., GORKA, A., HYDE, L. W., KIMAK, M., HALDER, I., DUCCI, F., FERRELL, R. E.,
 609 GOLDMAN, D. & MANUCK, S. B. 2009. Divergent effects of genetic variation in
 610 endocannabinoid signaling on human threat- and reward-related brain function. *Biol*
 611 *Psychiatry*, 66, 9-16.

612 HAYES, A. F. 2017. *Introduction to mediation, moderation, and conditional process analysis: A*
 613 *regression-based approach.* , Guilford Publications.

614 HEATHERTON, T. F., KOZLOWSKI, L. T., FRECKER, R. C. & FAGERSTRÖM, K. O. 1991. The
 615 Fagerström Test for Nicotine Dependence: a revision of the Fagerström Tolerance
 616 Questionnaire. *Br J Addict*, 86, 1119-27.

617 HENDERSHOT, C. S., WARDELL, J. D., MCPHEE, M. D. & RAMCHANDANI, V. A. 2017. A prospective
 618 study of genetic factors, human laboratory phenotypes, and heavy drinking in late
 619 adolescence. *Addict Biol*, 22, 1343-1354.

620 HINGSON, R. W., ZHA, W. & WHITE, A. M. 2017. Drinking Beyond the Binge Threshold: Predictors,
621 Consequences, and Changes in the U.S. *Am J Prev Med*, 52, 717-727.

622 KOOB, G. F. 2014. Neurocircuitry of alcohol addiction: synthesis from animal models. *In*:
623 SULLIVAN, E. V. & PFEFFERBAUM, A. (eds.) *Handbook of Clinical Neurology, Alcohol and*
624 *the Nervous System*. Elsevier.

625 KUNTSCHE, E., KNIBBE, R., GMEL, G. & ENGELS, R. 2005. Why do young people drink? A review of
626 drinking motives. *Clinical Psychology Rev*, 25, 841-861.

627 LINSENBARDT, D. N. & BOEHM, S. L., 2ND 2009. Agonism of the endocannabinoid system
628 modulates binge-like alcohol intake in male C57BL/6J mice: involvement of the posterior
629 ventral tegmental area. *Neuroscience*, 164, 424-34.

630 MALINEN, H. & HYYTIÄ, P. 2008. Ethanol self-administration is regulated by CB1 receptors in the
631 nucleus accumbens and ventral tegmental area in alcohol-preferring AA rats. *Alcohol Clin*
632 *Exp Res*, 32, 1976-83.

633 MANSOURI, E., NOBREGA, J. N., HILL, M. N., TYNDALE, R. F., LEE, F. S., HENDERSHOT, C. S., BEST,
634 L. M., DI CIANO, P., BALSEVICH, G., SLOAN, M. E., KISH, S. J., TONG, J., LE FOLL, B. &
635 BOILEAU, I. 2020. D3 dopamine receptors and a missense mutation of fatty acid amide
636 hydrolase linked in mouse and men: implication for addiction.
637 *Neuropsychopharmacology*, 45, 745-752.

638 MAYO, L. M., ASRATIAN, A., LINDÉ, J., HOLM, L., NÄTT, D., AUGIER, G., STENSSON, N.,
639 VECCHIARELLI, H. A., BALSEVICH, G., AUKEMA, R. J., GHAFOURI, B., SPAGNOLO, P. A., LEE,
640 F. S., HILL, M. N. & HEILIG, M. 2018. Protective effects of elevated anandamide on stress

641 and fear-related behaviors: translational evidence from humans and mice. *Mol*
642 *Psychiatry*.

643 MAYO, L. M., ASRATIAN, A., LINDE, J., MORENA, M., HAATAJA, R., HAMMAR, V., AUGIER, G., HILL,
644 M. N. & HEILIG, M. 2020. Elevated Anandamide, Enhanced Recall of Fear Extinction, and
645 Attenuated Stress Responses Following Inhibition of Fatty Acid Amide Hydrolase: A
646 Randomized, Controlled Experimental Medicine Trial. *Biol Psychiatry*, 87, 538-547.

647 MEZQUITA, L., STEWART, S. H., IBÁÑEZ, M. I., RUIPÉREZ, M. A., VILLA, H., MOYA, J. & ORTET, G.
648 2011. Drinking motives in clinical and general populations. *Eur Addict Res*, 17, 250-61.

649 MORENA, M., AUKEMA, R. J., LEITL, K. D., RASHID, A. J., VECCHIARELLI, H. A., JOSSELYN, S. A. &
650 HILL, M. N. 2019. Upregulation of Anandamide Hydrolysis in the Basolateral Complex of
651 Amygdala Reduces Fear Memory Expression and Indices of Stress and Anxiety. *J Neurosci*,
652 39, 1275-1292.

653 NATIONAL INSTITUTE ON ALCOHOL ABUSE AND ALCOHOLISM. 2020. *Alcohol Facts and Statistics*
654 [Online]. Available:
655 <https://www.niaaa.nih.gov/sites/default/files/AlcoholFactsAndStats.pdf> [Accessed June
656 15 2020].

657 NEIGHBORS, C., LEE, C. M., LEWIS, M. A., FOSSOS, N. & LARIMER, M. E. 2007. Are social norms
658 the best predictor of outcomes among heavy-drinking college students? *J Stud Alcohol*
659 *Drugs*, 68, 556-65.

660 PATEL, S., HILL, M. N., CHEER, J. F., WOTJAKI, C. T. & HOLMES, A. 2017. The endocannabinoid
661 system as a target for novel anxiolytic drugs. *Neuroscience and Biobehavioral Reviews*, 76,
662 56-66.

663 PAVON, F. J., SERRANO, A., STOUFFER, D. G., POLIS, I., ROBERTO, M., CRAVATT, B. F., MARTIN-
 664 FARDON, R., RODRIGUEZ DE FONSECA, F. & PARSONS, L. H. 2019. Ethanol-induced
 665 alterations in endocannabinoids and relevant neurotransmitters in the nucleus
 666 accumbens of fatty acid amide hydrolase knockout mice. *Addict Biol*, 24, 1204-1215.

667 POKORNY, A. D., MILLER, B. A. & KAPLAN, H. B. 1972. The brief MAST: a shortened version of the
 668 Michigan Alcoholism Screening Test. *Am J Psychiatry*, 129, 342-5.

669 RADEMACHER, D. J., MEIER, S. E., SHI, L., HO, W. S., JARRAHIAN, A. & HILLARD, C. J. 2008. Effects
 670 of acute and repeated restraint stress on endocannabinoid content in the amygdala,
 671 ventral striatum, and medial prefrontal cortex in mice. *Neuropharmacology*, 54, 108-16.

672 SAUNDERS, J. B., AASLAND, O. G., BABOR, T. F., DE LA FUENTE, J. R. & GRANT, M. 1993.
 673 Development of the Alcohol Use Disorders Identification Test (AUDIT): WHO Collaborative
 674 Project on Early Detection of Persons with Harmful Alcohol Consumption - II. *Addiction*,
 675 88, 791-804.

676 SCHERMA, M., MASIA, P., SATTA, V., FRATTA, W., FADDA, P. & TANDA, G. 2018. Brain activity of
 677 anandamide: a rewarding bliss? *Acta Pharmacol Sin*, 40, 309-23.

678 SIPE, J. C., CHIANG, K., GERBER, A. L., BEUTLER, E. & CRAVATT, B. F. 2002. A missense mutation in
 679 human fatty acid amide hydrolase associated with problem drug use. *Proceedings of the*
 680 *National Academy of Sciences*, 99, 8394-8399.

681 SIPE, J. C., SCOTT, T. M., MURRAY, S., HARISMENDY, O., SIMON, G. M., CRAVATT, B. F. & WAALEN,
 682 J. 2010. Biomarkers of endocannabinoid system activation in severe obesity. *PLoS One*, 5,
 683 e8792.

684 SLOAN, M. E., GOWIN, J. L., YAN, J., SCHWANDT, M. L., SPAGNOLO, P. A., SUN, H., HODGKINSON,
685 C. A., GOLDMAN, D. & RAMCHANDANI, V. A. 2017. Severity of alcohol dependence is
686 associated with fatty acid amide hydrolase Pro129Thr missense variant. *Addict Biol.*

687 SOBELL, L. & SOBELL, M. 1992. Timeline follow-back: a technique for assessing self-reported
688 alcohol consumption. In: LITTEN, R. & ALLEN, J. P. (eds.) *Measuring Alcohol Consumption:
689 Psychosocial and Biochemical Methods*. New York: Springer.

690 SPAGNOLO, P. A., RAMCHANDANI, V. A., SCHWANDT, M. L., KWAKO, L. E., GEORGE, D. T.,
691 HILLARD, C. J. & HEILIG, M. 2016. FAAH Gene Variation Moderates Stress Response and
692 Symptom Severity in Patients with Posttraumatic Stress Disorder and Comorbid Alcohol
693 Dependence. *Alcohol Clin Exp Res.*, 40, 2426-34.

694 TABACHNICK, B. G., FIDELL, L. S. & ULLMAN, J. B. 2007. *Using multivariate statistics*, Boston, MA,
695 Pearson.

696 TYNDALE, R. F., PAYNE, J. I., GERBER, A. L. & SIPE, J. C. 2007. The Fatty Acid Amide Hydrolase
697 C385A (P129T) Missense Variant in Cannabis Users: Studies of Drug Use and Dependence
698 in Caucasians. *American Journal of Medical Genetics Part B (Neuropsychiatric Genetics)*,
699 144B, 660-666.

700 VINOD, K. Y., MACCIONI, P., SALUD GARCIA-GUTIERREZ, M., FEMENIA, T., XIE, S., CARAI, M. A. M.,
701 MANZANARES, J., COOPER, T. B., HUNGUND, B. L. & COLOMBO, G. 2012. Innate difference
702 in the endocannabinoid signaling and its modulation by alcohol consumption in alcohol-
703 preferring sP rats. *Addict Biol*, 17, 65-75.

704 VINOD, K. Y., SANGUINO, E., YALAMANCHILI, R., MANZANARES, J. & HUNGUND, B. L. 2008.
705 Manipulation of fatty acid amide hydrolase functional activity alters sensitivity and
706 dependence to ethanol. *J Neurochem*, 104, 233-243.

707 ZHOU, Y., HUANG, T., LEE, F. & KREEK, M. J. 2016. Involvement of Endocannabinoids in Alcohol
708 "Binge" Drinking: Studies of Mice with Human Fatty Acid Amide Hydrolase Genetic
709 Variation and After CB1 Receptor Antagonists. *Alcohol Clin Exp Res*, 40, 467-73.

710 ZHOU, Y., SCHWARTZ, B. I., GIZA, J., GROSS, S. S., LEE, F. & KREEK, M. J. 2017. Blockade of alcohol
711 escalation and "relapse" drinking by pharmacological FAAH inhibition in male and female
712 C57BL/6J mice. *Psychopharmacology*, 234, 2955-2970.

713

714 **Table 1: Baseline demographics of participants**

Characteristics of the Study Sample	CC Genotype (n = 186)	AC/AA Genotype (n = 116)	p value	χ^2 / t value
Sex (Females/Males), n (%)	100 (54%)/86 (46%)	59 (51%)/57 (49%)	.623	0.24
Age, Years[†]	19.74 (1.18)	20.10 (1.3)	.012*	-2.53
Ethnicity (Caucasian/Non-caucasian), n (%)	98 (53%)/88 (47%)	55 (47%)/61 (53%)	.373	0.80
Currently Enrolled as a Student, n (%)	151 (81%)	85 (73%)	.328	2.23
Current Cigarette Smoker, n (%)	41 (22%)	29 (25%)	.560	0.34
Fagerstrom Test of Nicotine Dependence^{†,a}	1.12 (1.35)	1.33 (1.73)	.574	-0.56
<i>Self-reported Current Frequency of Cannabis Use, past 90 days</i>				
"Never", n (%)	62 (33%)	38 (33%)	.069	8.70
"Once or twice", n (%)	49 (26%)	23 (20%)		
"Monthly", n (%)	14 (8%)	12 (10%)		
"Weekly", n (%)	33 (18%)	13 (11%)		
"Daily or almost daily", n (%)	26 (14%)	29 (25%)		
Barratt Impulsivity Scale - Total[†]	64.6 (10.67) ^b	63.60 (10.80)	.411	0.82

715

Table 2: Results of the the ANCOVA analyses of covariate-adjusted mean differences between the genotype groups on the alcohol variables

	<i>FAAH</i> CC Genotype (n=186)		<i>FAAH</i> AC/AA Genotype (n=116)				
	Adjusted Mean†	SE	Adjusted Mean†	SE	<i>F</i>	<i>p</i>	partial η^2
<i>TLFB 90 Days</i>							
Number of drinking days	18.25	0.86	21.06	1.09	4.064	0.045*	0.013
Average Number of Drinks per Drinking Occasion	4.82	0.15	5.28	0.19	3.37	0.068	0.011
Number of Heavy Drinking Episodes	9.77	0.73	13.29	0.93	8.79	0.003**	0.029
<i>AUDIT</i>							
Total Score	9.84	0.41	11.17	0.52	4.05	0.045*	0.013
AUDIT-C	5.71	0.13	6.22	0.17	5.48	0.02*	0.018
<i>Drinking Motives Questionnaire</i>							
Coping Motives	10.59	0.33	12.12	0.42	8.13	0.005**	0.027
Enhancement Motives	16.14	0.36	17.16	0.46	3.07	0.081	0.01

Table Legends

Table 1 - †Values represented as mean (SD); * $p < 0.05$ for independent t-tests or Pearson's chi square tests conducted between genotype groups of the same parent study. ^abased on a subset of participants who reported regular nicotine use and completed the Fagerström Test of Nicotine Dependence ($n = 68$). ^bbased on a subset of participants who completed the Barratt Impulsivity Scale ($n = 278$).

Table 2 - Covariate-adjusted means, standard deviations, F tests, and effect size estimates (partial η^2) from the ANCOVAs comparing the AC/AA and C/C groups on outcomes from the 90-day Timeline Followback (TLFB), the Alcohol Use Disorders Identification Test (AUDIT) and the Drinking Motives Questionnaire. †means adjusted for covariates sex, race and age; * $p < 0.05$; ** $p < 0.01$.

Figure 1.

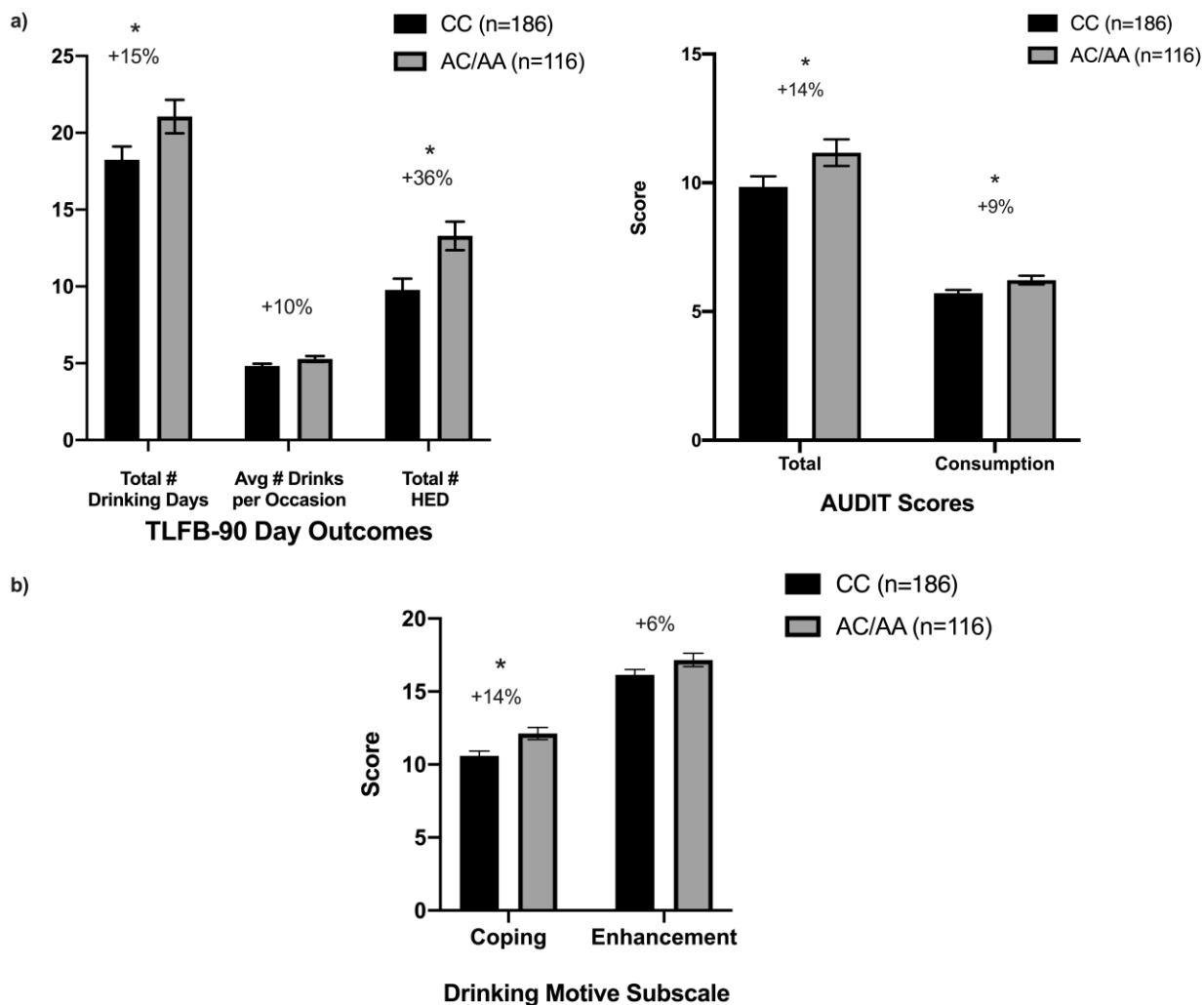


Figure 2.

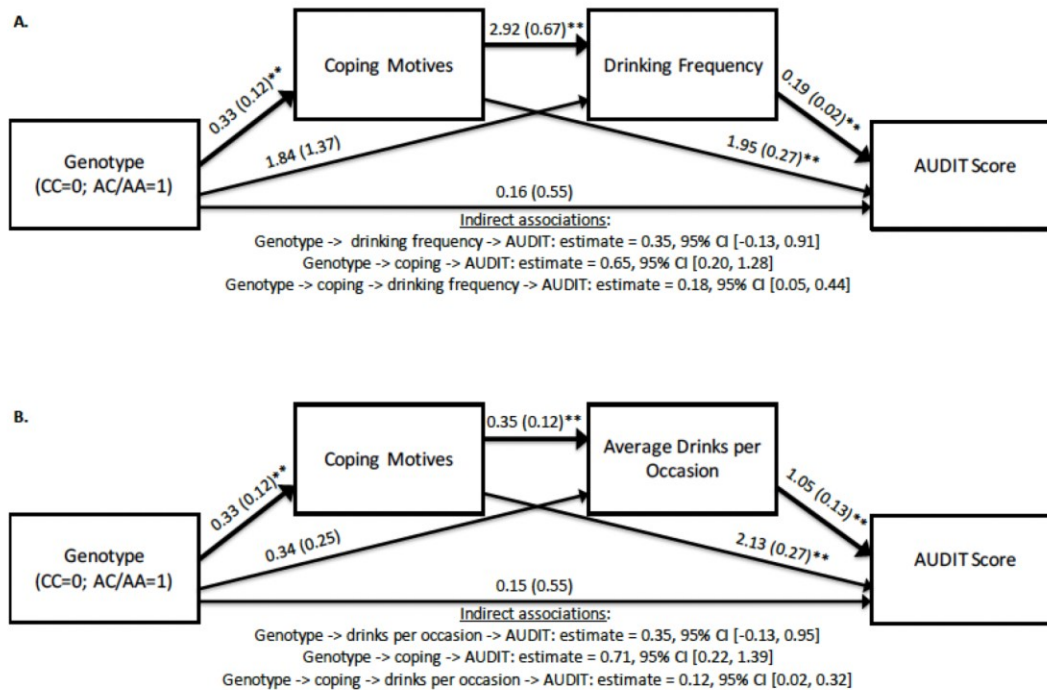


Figure Legends

Figure 1 – Comparison of covariate adjusted means from alcohol-related outcomes between FAAH C385A genotype groups (CC genotype vs AC/AA genotypes) using ANCOVAs with age, sex, and race as covariates. Alcohol consumption outcomes include the (a) Timeline Followback Method assessment of total number of drinking days (average number of drinks per drinking occasion and total number of heavy episodic drinking occasions in the past 90 days); (b) Alcohol Use Disorders Identification Test (AUDIT) total and consumption scores; and (c) Drinking Motives Subscale scores for coping and enhancement motives. * indicates a statistically significant difference ($p < 0.05$).

Figure 2 – Mediation models of indirect associations from genotype to coping motives to alcohol consumption (A. drinking frequency; B. average drinks per occasion; C. heavy episodic drinking frequency) to AUDIT score. Sex, race (white vs. non-white) and age are controlled in each model by regressing each outcome variable (coping motives, alcohol consumption and AUDIT) on each of these covariates. Estimates of the indirect associations contained within each model are shown below each path diagram with 95% bias-corrected confidence intervals (CI). * $p < 0.05$, ** $p < 0.01$.